

Electron microscopy of biogenic minerals: structure and sizes of uranium dioxide nanoparticles with Mn^{2+} impurities

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PACS 61.46.Df, 68.37.Lp, 87.17.-d

DOI 10.17586/2220-8054-2019-10-2-215-226

Methodological aspects of the extraction of structural and chemical information from transmission electron microscopy (TEM) of uranium dioxide (UO_2) biogenic nanoparticles are presented. Nanoparticles were formed via the bacterial reduction of water-soluble uranyl acetate with U (VI) in the presence of Mn^{2+} ions and cultures *Shewanella oneidensis* MR-1 in the medium. The particles of 1.2 – 3.5 nm in diameter and particle agglomerations were visualized in conventional TEM, high resolution TEM, scanning TEM modes. Their phase and chemical composition were investigated with electron diffraction, X-ray energy dispersive spectrometry and electron energy loss spectroscopy with high spatial resolution. Maintenance of the element balance helped to find the composition of the mixture of UO_2 and Mn acetates. The interpretation of TEM data and modeling allowed to propose the mechanism for the suppression of UO_2 particle growth and higher resistance to dissolution of smaller UO_2 particles with adsorbed Mn acetate compared to the larger pure particles.

Keywords: uranium dioxide nanoparticles, bacterial reduction, manganese impurity, transmission electron microscopy.

Received: 10 February 2019

Revised: 21 March 2019

1. Introduction

One of the primary reasons for investigating the biogenic minerals is to better understand the mechanism of interaction of living organisms and inorganic materials [1]. A direct involving of bacteria in the reduction of metals and metalloids (iron, chromium, uranium, selenium, tellurium and so on) is of considerable interest mainly because of the potential application in the treatment and bioremediation of uranium-contaminated groundwater. The basis of such treatment is the transformation of highly soluble U(VI) salts to sparingly soluble uranium (IV) oxide, uraninite. However, the strategy can be successful if the biogenic uraninite remains immobilized and does not easily oxidize [2]. Therefore, the structure and morphology of biogenic minerals, as well the impurity effect, have been and remain a subject of close attention with a view to addressing problems in fundamental and applied science and in technology.

The presence of different impurities has the potential to complicate the remediation process: change the rates of uranium transformations, influence the size and shapes of precipitated particles, enhance or inhibit dissolution and oxidation of the U(IV) oxide biogenic mineral. Manganese (Mn) is the 10th most abundant element in the Earth's crust and second only to iron as the most common heavy metal and Mn(II) is readily depleted from igneous and metamorphic rocks by interactions with surface water and groundwater being highly mobile in acidic aqueous systems [3]. Mn (II) presented in groundwater is considered a pollutant mainly because of their organoleptic properties [4]. Therefore, there is a request to develop environmentally friendly ways to remove Mn from water. In this regard, the combination of redox and adsorption processes is very attractive for removing two ions U(VI) and Mn(II) simultaneously.

Among all the methods available for the study of small particles, analytical transmission electron microscopy (TEM) offers possibilities for visualization in conventional TEM, high resolution TEM (HRTEM), scanning TEM (STEM) modes, investigation of phase (electron diffraction) and element composition (X-ray energy dispersive spectrometry, EDS), determination of chemical and oxidation state (electron energy loss spectroscopy, EELS) of individual particles of a few nm size, their agglomerations and mixtures of different phases with high spatial resolution. Severe requirements are imposed on thickness of TEM specimens. Clearly interpretable data can be obtained only for thin samples with a thickness of less than 50 nm and in some cases (for instance, using high-resolution methods) the thickness of the samples should not exceed 10 nm. The electron radiation effect on biogenic minerals can be a big issue. Small particles can change their shape and size, and such changes depend on both the time of irradiation and the intensity of the beam. The electron damage affects the structure and/or the chemistry of specimens when inelastic scattering (heating) breaks the chemical bonds (radiolysis) [5].

In this work, the particular interest was in identifying and characterizing the uranium-containing compound produced by metal-reducing bacteria in the presence of Mn impurities using TEM, electron diffraction, X-ray EDS and EELS. An additional objective is to provide insight methodological aspects, revisiting the TEM data coupling with electron diffraction simulation and structure modeling to propose the mechanism of higher resistance of smaller uraninite particles with adsorbed Mn acetate compared to the larger pure UO_2 particles.

2. Materials and methods

2.1. Cell cultures

Biogenic samples were prepared under anoxic conditions at École Polytechnique Fédérale de Lausanne (EPFL), Switzerland. All details of preparation of bacterial cultures (*Shewanella oneidensis* MR-1) for reduction of U(VI) and chemical experiments are given elsewhere [6–9].

2.2. Reduction of U(VI) in the presence of Mn impurities

A colony of *Shewanella oneidensis* MR-1 was grown in LB medium (water, peptide tryptone, yeast extract and NaCl). Reduction of U(VI) from uranyl acetate was carried out at pH 6.3 within 12 h for most of the conditions. The MnCl_2 solution was added to adjust to 0.1, 0.25, 0.5, 1.0, 2.5, 5.0 and 8.0 mM concentrations.

The cells and resulting nanoparticles were collected by centrifugation. The resulting pellet was washed with anoxic H_2O . Some part of samples was treated with 1 M NaOH to remove the biomass and 0.5 M NaHCO_3 to remove remaining adsorbed U(VI) as much as possible. Thus, the two main types of samples were investigated using TEM: samples containing cells and nanoparticles, and samples containing nanoparticles only. Mn^{2+} adsorbed on the particle surface and weakly bound to it was rinsed off using 25 ml of 20 mM MES (2-(N-morpholino) ethanesulfonic acid) buffer at pH 5. More details on specimen treatments and chemicals can be found elsewhere [7, 8].

2.3. Transmission electron microscopy

Samples for TEM study were prepared by drying the colloids of cells with particles on a carbon-coated copper grid (whole mount specimens) or as 50 – 80 nm slices of epoxy (Araldite, Epon or DER resin) with embedded cells and particles. In the latter case, bacterial specimens after reduction were fixed in 2.5 % glutaraldehyde and sequentially dehydrated in pure grade ethanol series (from 25 % to 100 %). The dehydrated cells with nanoparticles were collected by centrifugation and embedded in Araldite resin or Epon resin, then polymerized. The thin slices of polymerized resin were cut with an ultramicrotome using a diamond knife and then placed on copper grids with porous carbon films.

The structure and chemical composition of the particles were studied by conventional TEM, HRTEM, STEM, selected area electron diffraction (SAED), X-ray EDS and EELS in a CM300UT FEG (300 kV field emission gun, 0.65 nm spherical aberration, and 0.17 nm resolution at Scherzer defocus) and a Tecnai Osiris (200 kV high brightness XFEG, 0.24 nm point resolution, a-twin pole piece with Super-X EDX).

The TEM/HRTEM and EEL spectra images were processed with the Gatan Digital Micrograph software. The X-ray EDS microanalysis was performed in STEM mode and quantitative data were derived with the help of INCA (Oxford) and ESPRIT software (Bruker). Quantitative EDS analysis was carried out using the Cliff–Lorimer standard-less method at the nanoscale with thickness correction [5] and in this work were done assuming several possible thickness values from 0 to 60 nm.

The simulation and interpretation of SAED patterns and diffractograms (Fast Fourier transforms) of HRTEM images and structure modeling were performed with the JEMS software package [10] using the known crystal structures and atom positions, electron-optic parameters of the microscopes. The measurement errors in electron diffraction patterns and HRTEM images were about 5 % and 2 %, correspondingly.

3. Results and discussion

3.1. Imaging and composition of the biogenic minerals

The (S)TEM images showed cells coated by nanoparticles and nanoparticle agglomerations in the extracellular space (Figs. 1 and 2). The chemical and phase composition of biogenic minerals were derived from X-rays energy dispersive spectra (Fig. 1(b) and Fig. 2(c,d)) and SAED patterns (Fig. 1(d) and Fig. 2(b)) obtained from nanoparticles. The (S)TEM images in Fig. 1 obtained from the cells embedded in the resin show particles only in the extracellular space and the STEM image in the Fig. 2 taken from the whole mount sample demonstrates completely covered cells by U oxide particles together with the extracellular agglomerates.

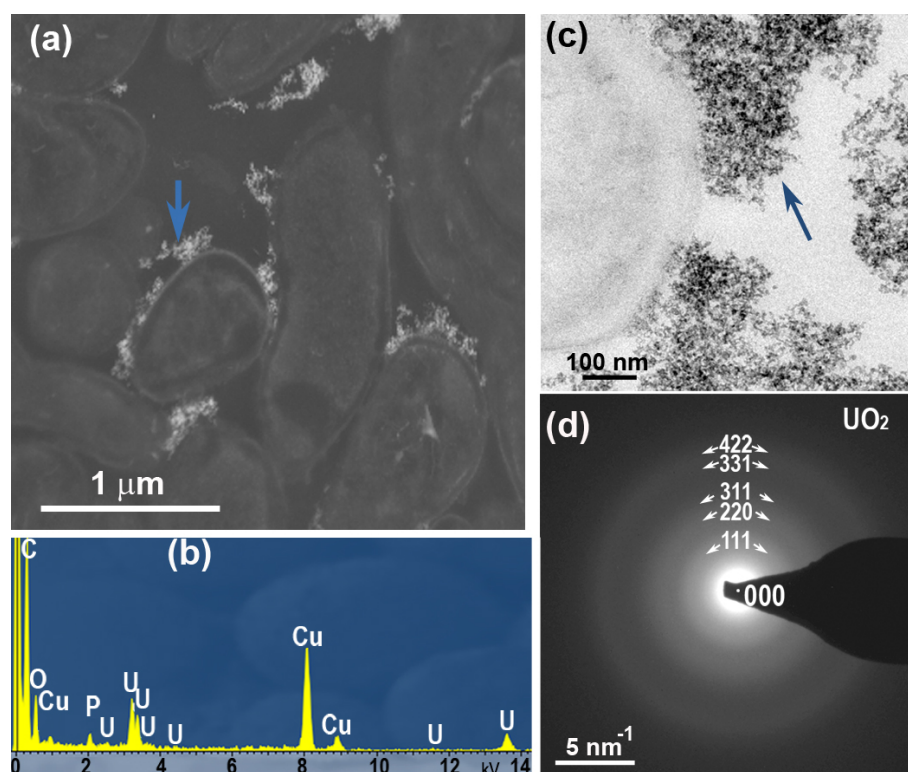


FIG. 1. STEM image of cells and UO_2 nanoparticles in an extracellular space embedded in araldite resin (a), X-ray EDS spectrum obtained from the particle agglomeration indicated by arrows (b), TEM image of one of cells and UO_2 nanoparticles (c) and the corresponding SAED pattern (d) obtained from the particle agglomeration indicated by an arrow in the TEM image.

The following elements were identified by X-ray EDS: U and O in the control sample without impurities (Fig. 1(b)), P comes from cells, C comes from cells and from the carbon support, Cu comes from the TEM grid. Chemical analysis of samples in which manganese chloride was added showed that manganese is mainly localized in agglomerates of UO_2 (Fig. 2(c, d)), where it is almost 10 times larger than on the surface of bacteria. The presence of chlorine from the MnCl_2 precursor was not detected in the dried samples because of complete washing out. Elements Fe, Co, Ni (from the microscope material) and Cu (from the supporting grid), which were not included in the samples were deconvoluted during quantitative microanalysis.

The phase composition of minerals is derived quite easily from the experimental SAED patterns which were compared with the simulated ones using JEMS for different U oxides (UO_3 , U_3O_8 , UO , UO_2 and U_4O_9). Crystallographic parameters for U oxides are listed in Table 1. Analysis showed that the simulated SAED patterns from the UO_2 phase with cubic structure (sp.gr. Fmm, $a = 0.544$ nm) had the best match with the experimental ones despite the broaden rings. The example of such matching is shown in Fig. 2(b). All other oxides (with the exception of UO) should have the reflections closer to the center of the SAED pattern in comparison with the UO_2 phase. However, no evidence has been obtained for the presence of reflections from other U oxides.

Quantitative data obtained from X-ray ED spectra showed a significant amount of oxygen which cannot be explained by the presence of UO_2 only. Thus, the EDS data led to the assumption that Mn oxides (MnO , MnO_2 , Mn_3O_4 and Mn_2O_3) or other compounds containing Mn and O are formed. Simulation of ring diffraction patterns from the mixture of UO_2 and some Mn oxides (not presented here) with the crystal parameters listed in Table 1 showed that even for the low content of Mn oxides in mixture of nanoparticles the SAED patterns must contain some reflections closer to the central beam. However the background of experimental SAED pattern corresponded only to the UO_2 ring pattern as well Mn oxide particles were never observed separately from UO_2 agglomerates. Precipitates of Mn oxides are characterized by morphology different from that of uranium oxide particles [26,27]. In addition, the concentrations of oxygen extracted from EDS data were high and the balance could not be reached with U and Mn oxides. Therefore, it was necessary to find another source of large amounts of oxygen. Among the other compounds, manganese acetates [28,29] could be the potential candidates.

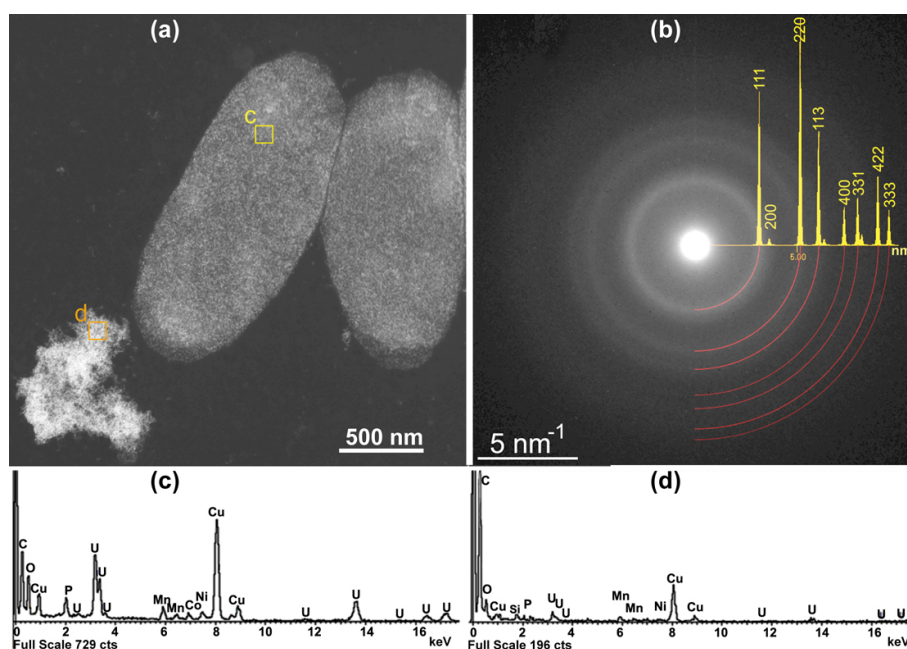


FIG. 2. TEM image of cells covered with UO₂ particles and UO₂ particles with Mn impurities in an extracellular space in a whole mount specimen (a), X-ray EDS spectra obtained from the particle agglomeration and the cell (b and c), SAED pattern obtained from the particle agglomerate (d), UO₂ phase with cubic structure derived using JEMS simulation of the SAED pattern (e)

Acetate ions, being a component of uranyl acetate and one of the major organic acid metabolites [30] after bacterial reduction of U (VI) can interact with Mn²⁺ ions and form hydrated manganese (II) acetates and adsorbed on the surface of UO₂ nanoparticles. It is known that simple mono-carboxylates, such as acetate, are able to generate complex extended structures due to the many accessible carboxylate bridging coordination modes [31]. Thus, the heterogeneous medium with UO₂ precipitates facilitates the formation and adsorption of Mn acetates. It also should be noted that in spite that, Mn acetate is considered as water soluble compound (solubility of tetrahydrate and anhydrous Mn acetate is 64.5 g/100 ml and 38.1 100 ml, respectively, at 50 °C) the solubility of MnCl₂ is much higher (88.5 g/100 ml at 40 °C) [32, 33] and therefore Cl-ions are much more easily washed out during rinsing with distilled water.

The formation of manganese acetate dihydrate or/and tetrahydrate corresponds well to elemental concentrations expressed in atomic % for the samples of assumed realistic thickness ranged from 10 to 60 nm. The plots (Fig. 3) show the actual oxygen concentrations in at.% depending on thickness for samples containing UO₂ particles with Mn impurities before and after treatment by the NaOH solution. The curve named “remaining oxygen” shows the remaining oxygen concentration of O after subtracting oxygen per uranium dioxide, phosphate group and possible silicon oxide. The lines named “MnAc₂ tetra”, “MnAc₂ dihydr”, “MnAc₂ anhydr” and “Mn hydroxide” show the oxygen concentration required to satisfy the corresponding stoichiometric formulas of Mn(CH₃COO)₂ · 4H₂O, Mn(CH₃COO)₂ · 2H₂O, Mn(CH₃COO)₂ and Mn(OH)₂. The intersection of these lines with the curve of “remaining oxygen” makes it possible to estimate the thickness of the sample.

Thus, the X-ray EDS data helped to maintain the balance of elements in accordance with the stoichiometric formulas to find the composition of a mixture of phases of UO₂ and Mn acetate tetra- or/and dihydrate within the experimental error.

The NaOH treatment of samples can lead to transformation of the part of Mn acetate into Mn hydroxide [34]. However, Mn acetate stays the main component of the adsorption layer and the UO₂ particle shell. This observation is consistent with a well-documented fact of the strong adsorption capacity of Mn²⁺ salts [35].

Figure 4 shows the structure models of UO₂ particles (core) coated by adsorbed manganese acetate tetrahydrate (Fig. 4(a)) on the 2.5-nm carbon film. The profiles of the simulated ring diffraction pattern (Fig. 4(b)) and the simulated ring diffraction patterns (Fig. 4(c)) evidence that the presence of Mn acetates in the given concentrations did not affect the positions of the peaks in electron diffraction patterns and show the good match with the experimental diffraction patterns.

TABLE 1. Crystallographic parameters of U and Mn oxides

Compound	Space group	Parameters [nm]	References
UO ₂	Cubic, Fm-3m	$a = 0.547$	[11]
UO	Cubic, Fm-3m	$a = 0.492$	[12]
UO ₃	Tetragonal, I41/am	$a = b = 0.6901,$ $c = 1.9975$	[13]
U ₃ O ₈	Hexagonal, P-3	$a = b = 0.6815,$ $c = 0.4136$	[14]
	Orthorhombic, C222	$a = 0.6704, b = 1.195,$ $c = 0.4142$	[15]
U ₄ O ₉	Cubic, I-43d	$a = 2.1805$	[16]
MnO	Cubic, Fm-3m	$a = 0.441$	[17]
	Cubic, Fm-3m	$a = 0.454$	[18]
MnO	Hexagonal, P63m	$a = 0.3372,$ $c = 0.5386$	[19]
Mn ₂ O ₃	Cub., Ia-3	$a = 0.941$	[20]
Mn ₃ O ₄	Tetragonal, I41/amd	$a = b = 0.5757,$ $c = 0.9424$	[21]
	Orthorhombic, Pbca	$a = 0.9412, b = 0.9418,$ $c = 0.9423$	[22]
MnO ₂	Tetr., P 42/mnm	$a = b = 0.4398,$ $c = 0.2873$	[23]
	Orthorhombic, Fddd	$a = 1.2207, b = 2.0154,$ $c = 0.2725$	[24]
	Orthorhombic, Pnma	$a = 0.9273, b = 0.2864,$ $c = 0.4522$	[25]

The presence of Mn and high content of oxygen in particle agglomeration and nearly the same U/Mn ratio in NaOH treated and untreated samples showed that the tendency of UO₂ particles to strongly agglomerate was preserved as well the major part of manganese acetate was not dissolved. Therefore, the process of biogenic removal of U⁶⁺ from contaminated water can be efficient also for the simultaneous removal of Mn²⁺ ions.

3.2. EELS characterization

Electron energy loss spectroscopy (EELS) was applied to ascertain if mixed chemical states of uranium (for instance, traces of the remaining U⁶⁺) and manganese can be determined after the bacterial reduction.

A typical EEL spectrum for UO₂ (U⁴⁺) obtained at room temperature with two U-M_{5,4} edges (3552 eV and 3728 eV, correspondingly) is present in Fig. 5(a). Applying the comparison of the experimental EEL spectra from the biogenic mineral and synthetic UO₂ standard namely M5/M4 white-line ratios [36] led to the conclusion that U⁶⁺ changed to U⁴⁺ and UO₂ particles formed.

However, the efforts to determine the valence state of the impurity Mn ion in the presence of large quantity of UO₂ failed. Fig. 5(b) shows the weak U-N_{7,6} edges between 350 and 400 eV and the O-K edge corresponding to the UO₂ phase while Mn-L_{3,2} lines were detected only for the specimens with the highest MnCl₂ concentration (8 mM) in the solution. We did not succeed to record clear Mn edges in the specimens with lowest concentration of Mn ion (when the UO₂ particles had the biggest size) [8]. Mn-L_{3,2} lines did not appear in the EEL spectrum due two possible reasons: a very low concentration of Mn ions (0.1 mM in solution) absorbed onto a thick UO₂ particle agglomeration. The increase in Mn concentration allowed detection of the Mn-L_{3,2} edges. However, EELS

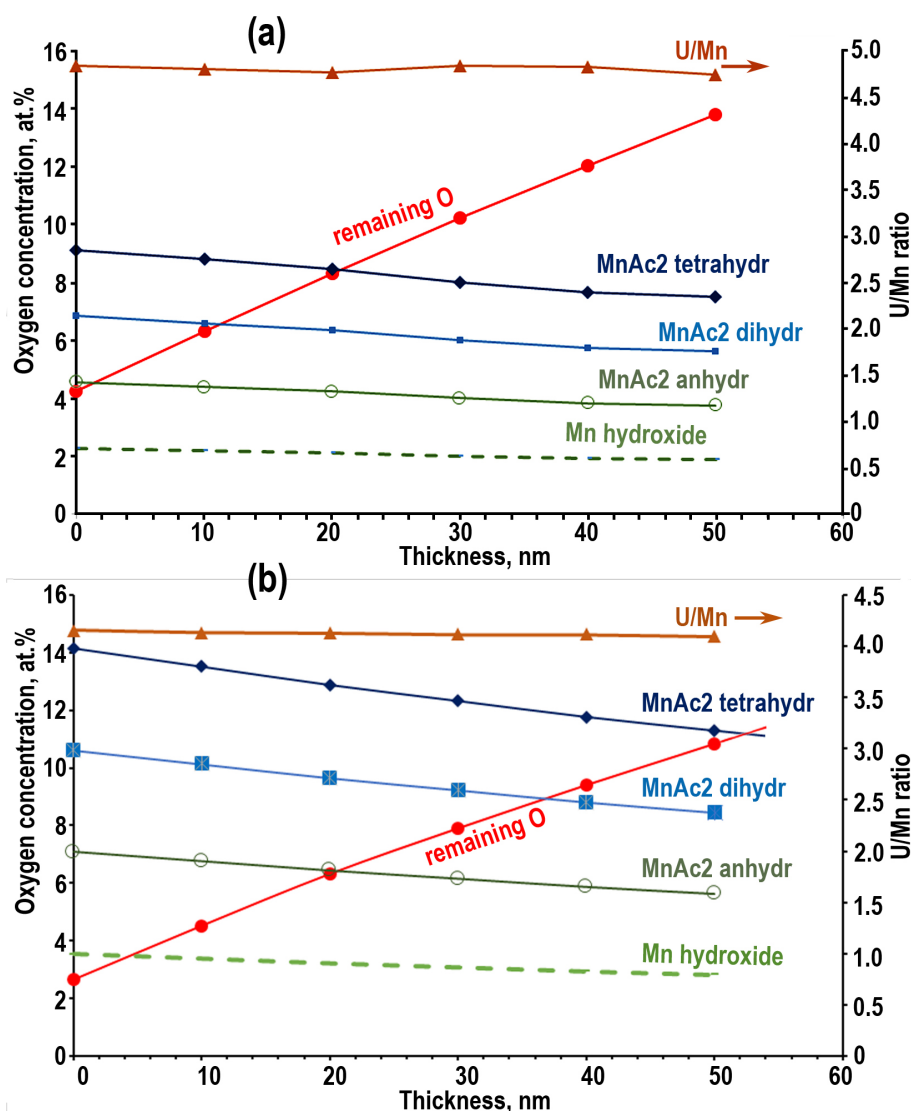


FIG. 3. Oxygen concentration values calculated using absorption correction method in the thickness range from 0 to 60 nm for untreated samples (a) and for NaOH treated samples (b). The ratio U/Mn is nearly constant and equal ≈ 4.7 for untreated samples (a) and ≈ 4.2 for NaOH treated ones (b)

measurements did not give an unambiguous answer to the question of the valence of Mn ions, which, again may be due to the insufficient Mn content and relatively thick UO_2 agglomerated specimens.

The significant effect of sample thickness and carbon on the U oxide spectrum in the low-loss region 0 – 60 eV [37] is shown in Fig. 5(c, d). Spectrum in Fig. 5(c) was obtained from the thick part of the UO_2 particle agglomeration on the carbon support while spectrum in Fig. 5(d) shows the spectrum from the thin part of the agglomeration over a hole in the carbon supporting film.

In contrast to some reports that the electron beam tends to change elemental oxidation state of U during EEL spectra acquisitions [37] we did not observe phase transformations and the appearance of other U phases in detectable quantities.

3.3. Determination of the particle size

One of the main objectives of this study was to determine whether the UO_2 particle sizes change with the addition of Mn impurities. Several techniques were applied to estimate the size of biogenic UO_2 particles. The use of electron diffraction is an indirect way to determine the average size of diffracting grains while (S)TEM imaging

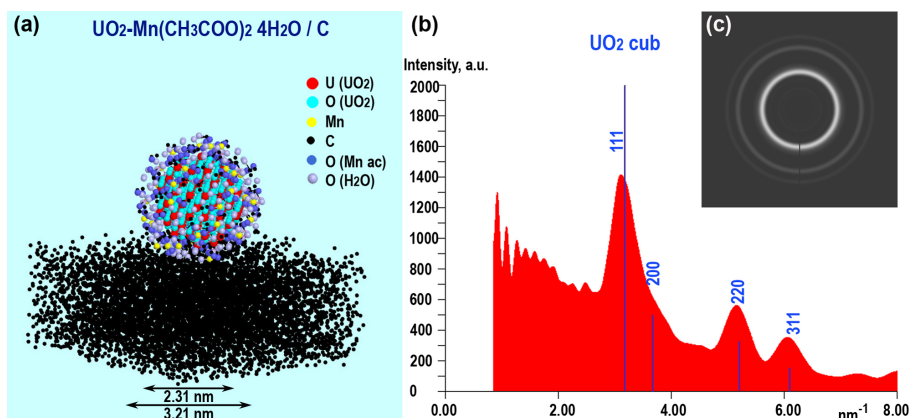


FIG. 4. Structural model of the UO_2 particle of 2.31 nm in diameter (core) coated by $\text{Mn}(\text{CH}_3\text{CO}_2)_2$ tetrahydrate (shell) of 0.9 nm thick on the carbon film of 2.5 nm thick (a), the simulated intensity profile of the diffraction rings with indicated UO_2 reflections (b), the simulated ring diffraction patterns from the $\text{Mn}(\text{CH}_3\text{CO}_2)_2$ tetrahydrate coated UO_2 particles (c)

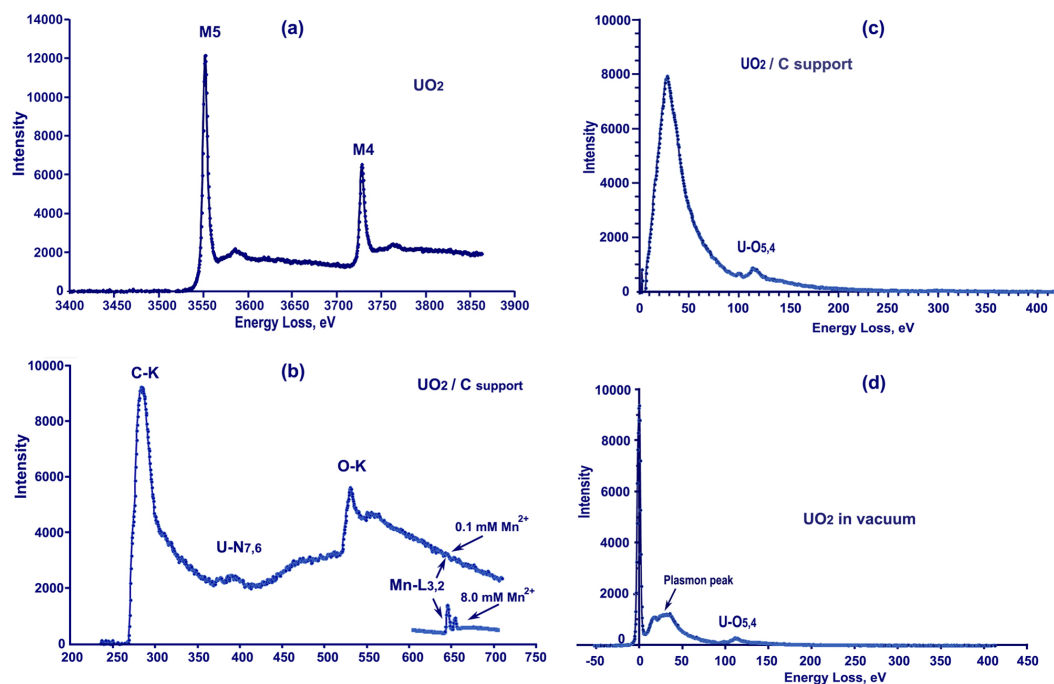


FIG. 5. $\text{U-M}_{4,5}$ EEL spectrum after background subtraction for uranium dioxide (a), $\text{U-N}_{7,6}$, O-K, C-K and Mn-L_{3,2} (inset) EEL spectrum after background subtraction for uranium dioxide and MnCl_2 on a carbon support (b), EEL spectrum obtained from the thick sample on the carbon support (c), EEL spectrum obtained from the thin sample over a hole in the carbon film in vacuum (d)

offers the direct way to measure sizes and particle size distributions. However, measurements using both methods are difficult due to the small particle size and their heavy agglomeration.

The average size of particles can be found by measuring the broadened electron diffraction rings and applying the Scherrer formula. However, if the sizes of the crystal nanoparticles are less than 4 nm, the relative error of the Scherrer formula increases significantly [38]. In this work, the simulation of powder (ring) diffraction patterns for particles with sizes in the range from 3.0 nm to 1.5 nm was performed. Afterwards the profiles of 111 + 200 rings in experimental electron diffraction patterns were compared with the simulated ones. The 111 reflection was chosen because it is the strongest reflection for UO_2 cubic phase and the corresponding ring is the most intense in

SAED patterns. The other 200 ring is quite close to the 111 ring and both rings start overlapping due to size effect broadening. Therefore, the intensity profile from both reflections were analyzed.

Figure 6 shows the TEM image (Fig. (a)), the corresponding SAED pattern (Fig. 6(b)), the HRTEM image of UO_2 particles (Fig. 6(c)) and the HRTEM FFT (Fig. 6(d)) obtained from the specimen containing 0.5 mM Mn^{2+} . A similar observation is present in Fig. 6(e) for UO_2 particles from the specimen containing 5 mM Mn^{2+} . The intensities of the SAED rings (Fig. 6(f)) decrease with the Mn concentrations as well the sizes of particles on the HRTEM image (Fig. 6(g)) also decrease and the rings in the FFTs become practically indistinguishable (Fig. 6(h)).

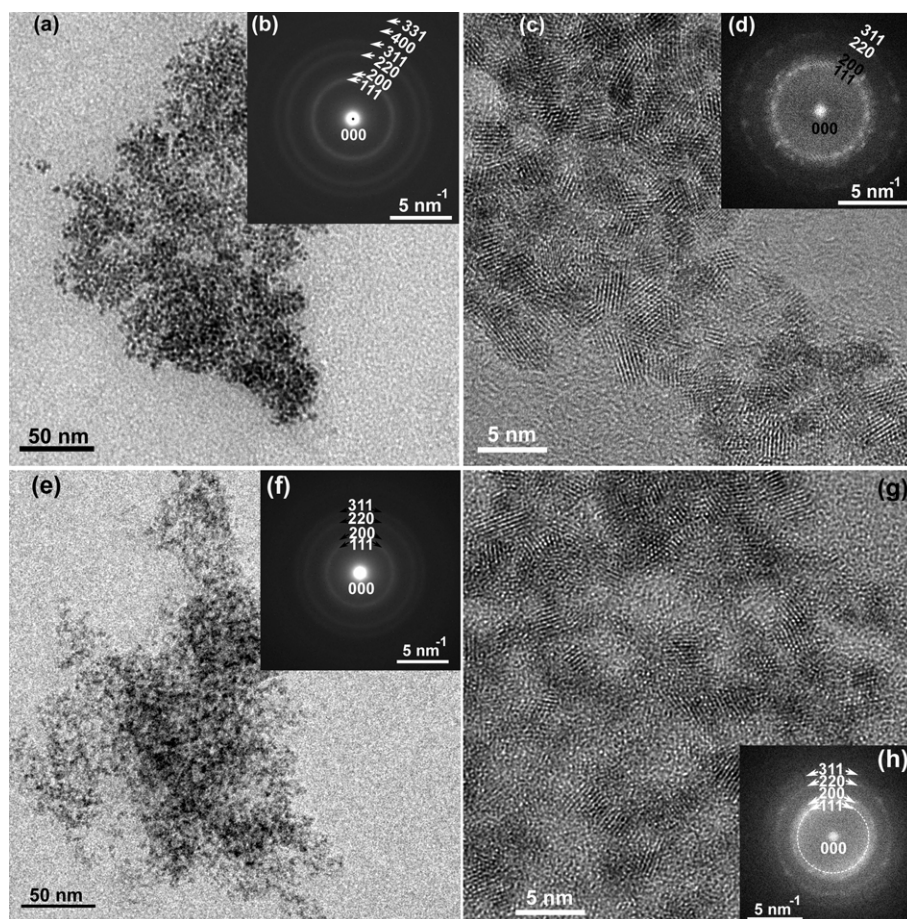


FIG. 6. TEM image of an agglomeration of UO_2 particles with the low concentration of Mn^{2+} impurities (a), ring SAED pattern (b), HRTEM image of UO_2 particles (c), FFT of the HRTEM image of UO_2 particles (d); TEM image of an agglomeration of UO_2 particles with the high concentration of Mn^{2+} impurities (e), ring SAED pattern (f), HRTEM image of UO_2 particles (g), FFT of the HRTEM image of UO_2 particles (h)

Analysis of the intensity profiles from both SAED patterns is shown in Figs. 7 and 8. The corresponding profiles (Figs. 7(b) and 8(b)) were recorded along the directions indicated in the SAED patterns (Figs. 7(a) and 8(a)), the peaks, including the 111 and 200 reflections after polynomial background subtraction, are shown in Figs. 7(c) and 8(c). Finally, fitting the 111 + 200 simulated peaks (dash lines in Figs. 7(d) and 8(d)) to the experimental ones was obtained for particles with the average size 2.4 nm and 1.9 nm, respectively.

The average particle size (the mode diameter) was also derived from the size distributions obtained from the HRTEM images (Fig. 9(a, c)). In order to recognize the edges of small particles (specimen with higher Mn concentration) the filtered HRTEM image was analyzed (Fig. 9(d)). The smaller the size of the particles, the farther their size distribution from the Gaussian shape (Fig. 9(b, d)), approaching the right skewed lognormal distribution.

The results reported earlier [39] showed that the fastest re-oxidation is characteristic for the smallest (≈ 1.5 nm) biogenic UO_2 particles. Adsorption of Mn acetates on the surface of UO_2 particles and formation of the shell can explain the increase of their stability observed in [6].

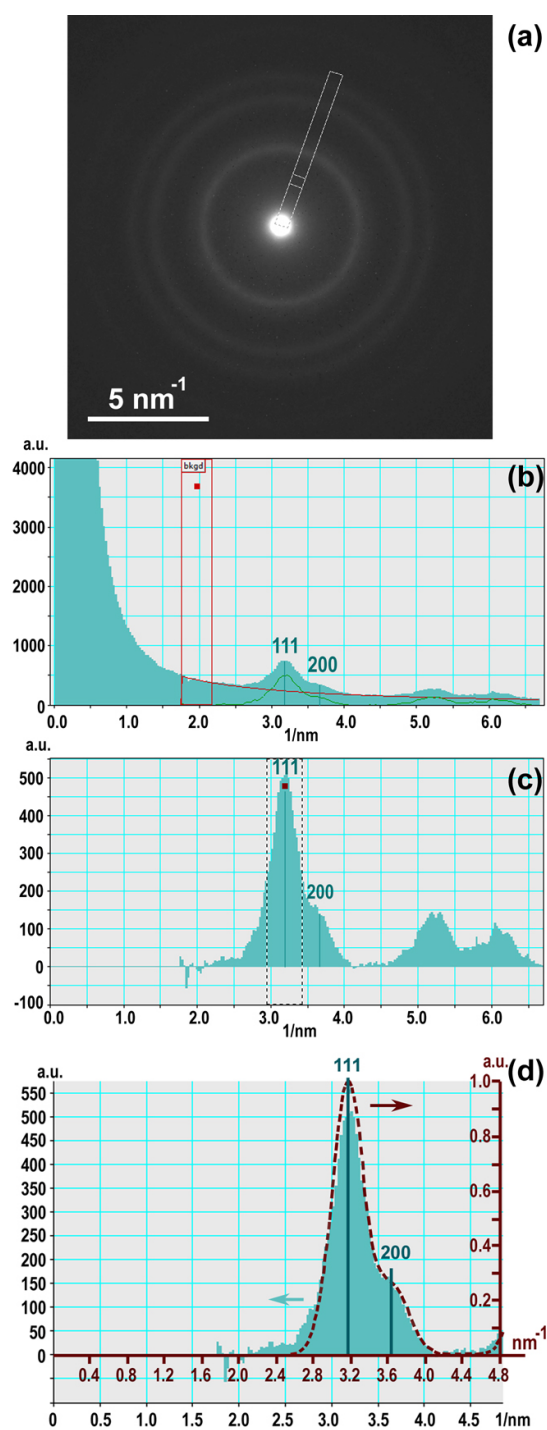


FIG. 7. Ring SAED patterns obtained from UO_2 particles with low Mn^{2+} concentration (a), the corresponding profiles of the SAED rings (b), signal intensity of 111 and 200 reflections after background subtraction (c), fitting the calculated profile of 111 + 200 reflection to the experimental intensity profile (d)

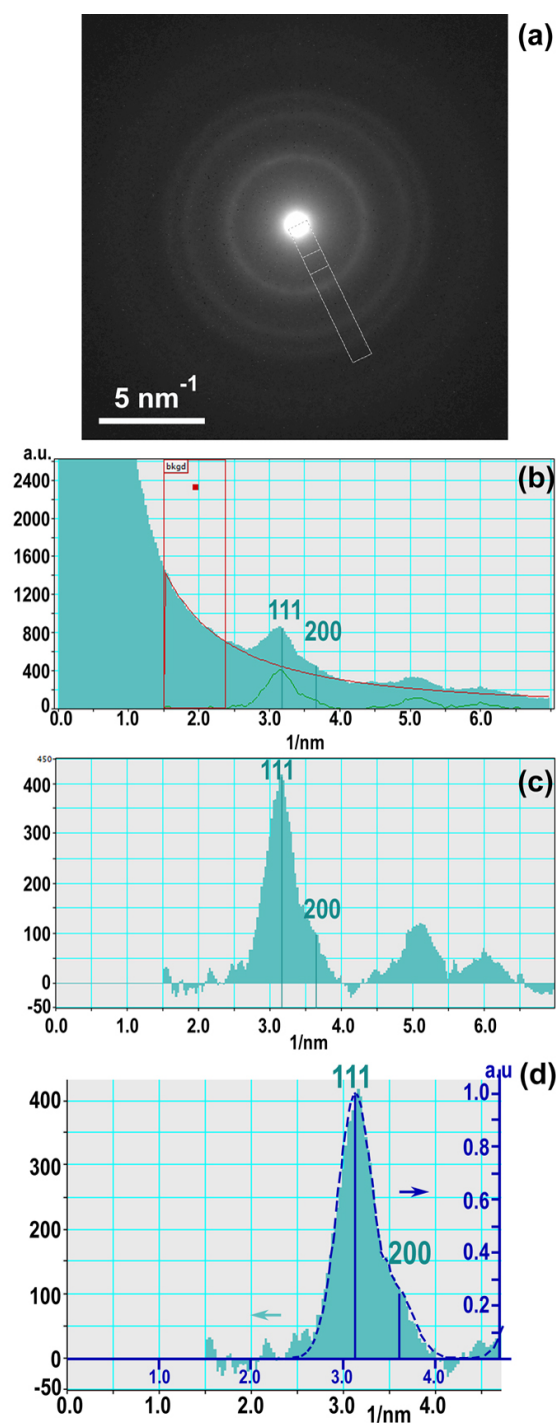


FIG. 8. Ring SAED patterns obtained from UO_2 particles with high Mn^{2+} concentration (a), the corresponding profiles of the SAED rings (b), signal intensity of 111 and 200 reflections after background subtraction (c), fitting the calculated profile of 111 + 200 reflection to the experimental intensity profile (d)

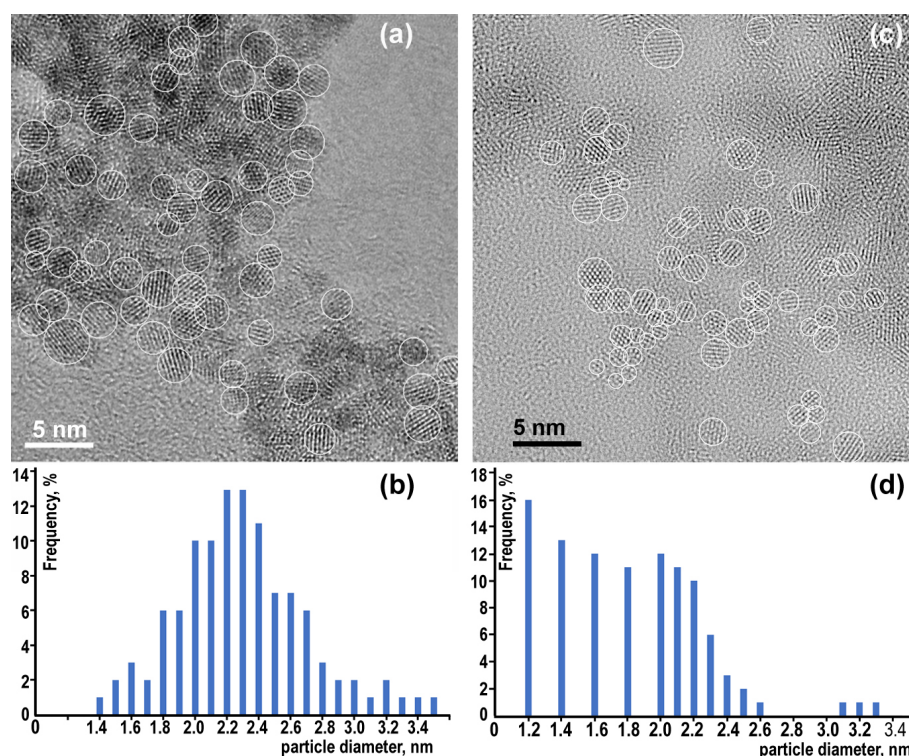


FIG. 9. HRTEM image of counted UO_2 particles with measured sizes with low (a) and high Mn^{2+} concentration (c) and the corresponding histograms (b, d). Histograms were obtained for 100 – 120 particles from several HRTEM images

Burgos et al. [39] reported that estimation of particle sizes with EXAFS (extended X-ray absorption fine-structure spectroscopy) could provide values of about 1.2 nm in diameter and such particles too small to be visualized by TEM what is not entirely true. Modeling the small UO_2 particles and the corresponding simulation of the HRTEM images (Fig. 10) showed that it is possible to visualize particles even smaller than 1 nm using the microscopes without aberration corrections.

The fact is that such small particles are unlikely to survive in solution having the sizes lower than the possible critical nucleus. Simulation of small UO_2 nanoparticles with non-integer number of cells showed a discrete set of sizes and the smallest particles found in HRTEM images had about 1.1 – 1.2 nm in diameter (Fig. 9(a, c)) what correspond to the model with diameter of 1.16 nm. And the next smallest size is about 0.76 nm. Such particles with 4 uranium atoms surrounded by 16 oxygen atoms are unlikely to stay independently.

3.4. Effect of the electron beam: Sintering of particles

The efforts to determine the particle size distribution met the problem of the electron irradiation induced sintering. The particle sizes increased with irradiation time or/and beam intensity. Diffractograms from HRTEM images obtained during different exposure time demonstrate the increase of particle size after electron irradiation for 40 s (Fig. 11).

Sintering the UO_2 nanoparticles did not lead to phase transformation and after 5 min of irradiation only 5.0 – 9.0 nm UO_2 irregular particles were found. Mn acetate shells also did not prevent sintering and did not introduce significant effect on the final sizes and phase composition of particles. However, the amount of oxygen lowered due to radiolysis and breaking bounds in Mn acetate structure since the electron diffraction patterns showed the UO_2 cubic structure.

Therefore, low dose and reasonably short exposure time should be used if the final target is to image nanoparticles and estimate their original sizes.

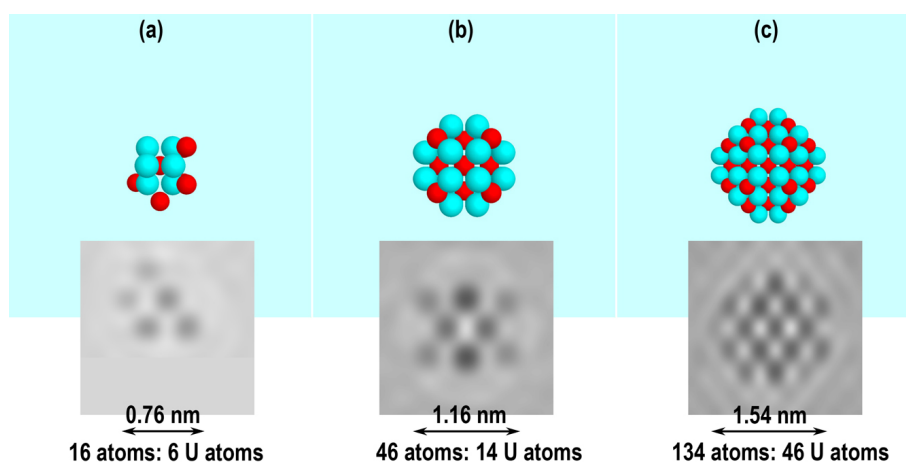


FIG. 10. The structural models of UO_2 particles are built with the help of JEMS (Stadelman, JEMS 2016) with different amount of unit cells embedded in boxes with sizes $2 \times 3 \times 3$ unit cells (a), $3 \times 3 \times 3$ unit cells (b) and $4 \times 4 \times 4$ unit cells (c) with particle diameters 0.76, 1.16 and 1.54 nm respectively. The HRTEM image simulation given under the corresponding model was performed using weak phase object assumption and the following imaging conditions: accelerating voltage 200 kV, spherical aberration coefficient $c_s = 1.4$ mm, chromatic aberration coefficient $c_c = 1.8$ mm, Scherzer defocus 72.5 nm, energy spread 0.8 eV

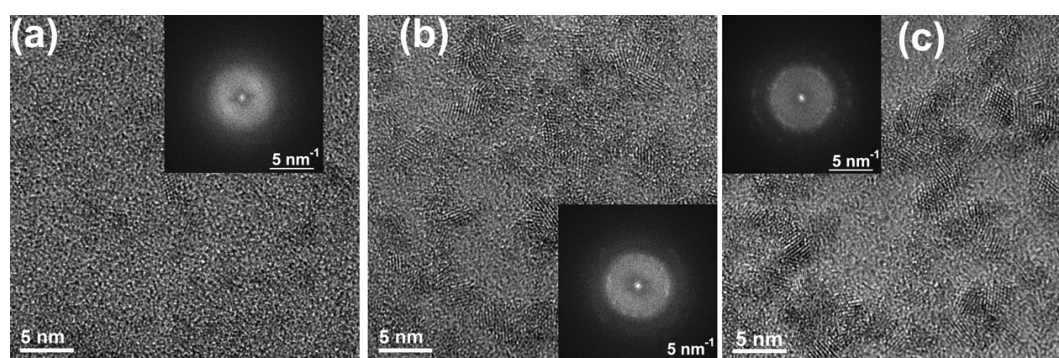


FIG. 11. HRTEM images and FFTs (insets) of UO_2 nanoparticles precipitated in solutions with different concentrations of Mn^{2+} ions: 8.0 mM (a), 1.0 mM (b), 0.1 mM (c)

4. Conclusions

Transmission electron microscopy analyses allowed study of the morphology and structure of biogenic nanoparticles formed during reduction of U(VI) from uranyl acetate using *S. oneidensis* MR-1 in the presence of Mn^{2+} impurities. UO_2 nanoparticles cover the surface of cells and are agglomerated in extracellular space. Their sizes decrease with increase of concentration of impurity Mn^{2+} ions in the solutions. Quantitative X-ray EDS microanalysis showed that Mn acetate tetra- or dihydrate formed and strongly adsorbed on the particle surface, which may explain suppression of UO_2 particle growth and the increase in particle stability and dissolution resistance. The UO_2 particles have a strong tendency to sinter under the electron beam and their size increases markedly while the presence of Mn acetate shell has no effect on the sintering process. Some elongated particles of approximately 8.0 – 9.0 nm long with irregular shape formed during sintering. The UO_2 phase is quite stable under electron beam and does not undergo phase transitions. The process of biogenic removal of U^{6+} from contaminated water can be efficient also for the simultaneous removal of Mn^{2+} ions.

Acknowledgements

All samples were prepared at École Polytechnique Fédérale de Lausanne (EPFL). The help of Prof. Philippe Buffat in discussions of results is acknowledged. This work was also supported by the Ministry of Science and Higher Education of the Russian Federation.

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